Product Information

Cat.No.	0566-CSC-C6655J
Product Name	SUIT-2
Description	human pancreatic cancer cell line establesh from liver metastasis
Recommended Medium	RPMI-1640 + 10% FBS
Morphology	epithelial-like
Instruction for Culturing	**Subculture:** split culture prior to confluence 1:2 to 1:5 every 2-5 days using trypsin/EDTA treatment; seeding at 3-5*10^5 cells/ml **Doubling time:** ~33 hours **Incubation:** at 37 °C with 5% CO_2 **Storage:** frozen with 70% RPMI-1640, 20% FBS,10% DMSO
Quality Control	**Mycoplasma:** negative in microbiological culture, PCR assays **Viruses:** PCR: EBV -, HBV -, HCV -, HHV-8 -, HIV-1 -, HIV-2 -, HTLV-I/II -, MLV -, SMRV -
Storage	LN_2.
Shipping	Dry Ice.
Protocol for Thawing Frozen Cells	1. Cells should be stored in liquid nitrogen. **DO NOT store cells at -80°C.** The cells are extremely temperature-sensitive and should be transferred to liquid nitrogen immediately upon arrival. Cells should be transported on dry ice or in a liquid nitrogen container. When transporting the cells on dry ice make sure that the vials are completely covered. 2. Read the cell line data sheet to establish specific requirements for your cell line. 3. Collect an ampoule of cells from liquid nitrogen storage wearing appropriate personal protective equipment and transfer to the laboratory in a container of liquid nitrogen or on dry ice. It is important to handle the ampoules with care: on rare occasions ampoules may explode on warming due to expansion of trapped residual liquid nitrogen. 4. Remove the cryovial containing the frozen cells from liquid nitrogen storage and immediately place it into a 37°C water bath. Quickly thaw the cells in less than 1 minute by gently swirling the vial in the 37°C water bath. **Note: It is important to thaw rapidly to minimize any damage to the cell membranes. Do not totally immerse the ampoule as this may increase the risk of contamination.** 5. Wipe ampoule with 70% alcohol prior to opening. 6. Pipette the whole content of the ampoule into a sterile tube (e.g. 15 ml capacity) with 5ml pre-warmed complete medium. 8. Centrifuge the cell suspension at approximately 230 × g for 5–10 minutes. 9. After the centrifugation, check the clarity of supernatant and visibility of a complete pellet. 10. Aseptically decant the supernatant without disturbing the cell pellet and resuspend the cell pellet in a volume of complete medium appropriate for counting the cells. **Count cells using a hemacytometer.**

Note: Do not seed the cryopreserved cells directly into your assay plates.

11. Seed the cells into the appropriate culture vessel to achieve **the recommended seeding density of viable cells**.

Note: NEVER CAN FROZEN CELLS BE KEPT AT -20 °C.

Gentaur Molecular Products BVBA
Address: Voortstraat 49, 1910 Kampenhout, Belgium
T: 0032 16 58 90 45 | **E:** info@gentaur.com
Websites: www.gentaur.com | www.maxanim.com